Prevalence of pathogens and microbiological quality of milk marketed in the region of the Recôncavo from Bahia, Brazil

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Abstract— Current study evaluates the sanitary quality and the presence of pathogens of crudeand processed bovine milk samples. Fifty-five samples from five municipalities in the Recôncavo da Bahia region were analyzed between May and July 2015. Psichrotrophic, mesophilic and thermophile rates were counted by depth plating method in standard agar; HiCrome ™ selective ECC was employed to quantify total coliforms and Escherichia coli. Escherichia coli 0157: H7 was identified with fast method Singlepath®- E. coli 0157. Staphylococcus aureuspopulation was estimated with Petrifilm ™ plates and Listeria monocytogeneswas identified by Singlepath L'Mono® kit. There was a greater contamination and presence of pathogens in the raw milk when compared to processed milk. However, total coliformswere detected in 14.28 %,Escherichia coliin 7.14 % and Escherichia coli 0157: H7 in 2.04 % of processed milk. The enforcement by authorities against the illegal sale of raw milk and the monitoring of steps in milk production up to marketing should be mandatory.

Keywords—sanitary conditions, dairy industry, food safety.

I. INTRODUCTION

Brazil currently ranks fourth in milk production worldwide (USDA, 2015), with more than 24 billion liters of raw milk in 2015. The Brazilian state of Minas Geraishad the greatest milk production, with 6,439,650liters, whilst, during the same year, the state of Bahia produced 332,449,000liters, featuring the greatest milk producers in the northeastern region of Brazil in 2015 (Instituto Brasileiro de Geografia e Estatística, 2016).

However, deficient hygiene conditions during milking, handling and conservation are the main factors for decrease in milk quality produced in Brazil (Menezes et al., 2015), damaging the country's economic development in milk production.

Although Brazilian law forbids selling of milk *in natura*, it is still common practice in Brazil, associated with cultural, regional and social factors (Bersot et al., 2010). Such practice may be harmful to consumer's health since milk is a good vector of pathogenic microorganisms which cause Food-Transmitted Diseases (FTD) (Claeys et al., 2013).

Dairies use thermal processes that reduce initial microbial load of raw milk, such as pasteurization and *Ultra High Temperature (UHT)*, to commercialize safer food with regard to sanitary hygiene and shelf time increase.

Further, several studies have revealed the bad quality of milk produced in Brazil (Tamanini et al., 2011; Pereira et al., 2013; Weschenfelder et al., 2016), affected not merely by prime matter used but by lack of post-processing health control, especially during bottling and packaging. Prevention against milk contamination during milking and storing is mandatory to reduce microbial multiplication and to produce quality milk products (Salvador, 2012).

Since milk is a high economic asset in Brazil and its intake may be a health risk for the population due to microbial multiplication, current paper assesses raw and UHT milk quality sold in the towns of the Recôncavo da Bahia region. The study may also be a warning for authorities for drastic control on milk producing farms and in the region's market.

II. MATERIAL AND METHOD

Fifty-five bovine milk samples were retrieved and analyzed between May and June 2015: four samples of untreated milk and seven whole UHT milk samples of different trademarks, from each municipality. Samples were purchased in five municipalities in the Recôncavo da Bahia region, namely, Cruz das Almas, Sapeaçu, Governador Mangabeira, Muritiba and São Felipe. Physical integrity, air tightness, packaging and lots verified during UHT milk sample collection were the selection criteria.

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Samples were maintained in isothermal boxes with recyclable ice and immediately transported to the Laboratory for Food and Water Analytic Investigation of Agrarian, Environmental and Biological Sciences of the Universidade Federal do Recôncavo da Bahia, Cruz das Almas BA Brazil..

Packages for raw and UHT milk were initially washed with water and neutral detergent, dried with white disposable paper, hygienized with ethanol 70% and homogenized (shaken 25 times). UHT milk samples were incubated at 35-37°C for seven days, following Brazilian legislation (1997, 2001).

Psichrotrophic, mesophyll and thermophile microorganisms were counted by pour plate technique in Plate Count Agar (PCA) medium (APHA 1.05463.0500). Further, 1 mL of each dilution was transferred to sterilized petri plates with 25 mL agar previously combined and warmed at 43-45°C. After homogenization and solidification, the plates were incubated in a buffer at 7°C for 10 days; 35°C for 48h; 50°C for 48h for psichrotrophic, mesophyll and thermophile microorganisms, respectively.

Coliforms were counted in medium Agar HiCrome™ Seletive ECC - M1294 Himedia® (ISO 9001:2008). Methodology complied with manufacturer's instructions. Colonies ranging between salmon and red color were typical colonies of total coliforms, while dark blue to violet revealed *Escherichia coli* colonies.

Staphylococcus aureus populations were calculated by fast method on plates (3M Company) PetrifilmTM (AOAC 2003.11), following manufacturer's instructions. Colonies of a red-violet color were typical of *S.aureus*.

E. coli O157:H7 analysis initially comprised a warming stage; the samples were then added to the selective enrichment broth MTSB Novobiocin, MERCKTM, and incubated at 35°C - 37°C for 18-24 hours (Ahmed & Shimamoto, 2014).

Species of *E. coli* O157:H7 was identified by fast immunological scanning Singlepath®- *E. coli* O157, MERCKTM (AOAC 010407), following manufacturer's instructions. Test apparatus was incubated at room temperature and result was given 20 min after the sample was applied to the apparatus. A negative result for strain *E. coli* O157:H7 occurred when only a red line appeared within the control zone (C); result was positive when red line appeared in tests (T) and control (C).

Listeria monocytogeneswas identified by fast kit Singlepath L'mono®, MERCKTM (1.04148.0001). Milk samples were previously added to Brain and Heart Infusion (BHI) broth and incubated at 29-30°C for 24 hours, for selective enrichment. The presence of *Listeria monocytogenes* in kits with red line in the test zone (T) and in the control zone (C) of the apparatus was positive, but negative when no line occurred in the test zone (T), although it appeared clearly in the control zone (C).

Mean number of colonies in all plates was multiplied by the respective dilution factor and results were given in log CFU/mL (Brasil, 2003). Moreover, statistical analysis was undertaken by SPSS 17. Averages and standard deviation of the microorganisms were calculated with descriptive analysis.

Means were compared by Student's t test for independent samples and evaluated whether there were any differences in the quantity of microorganisms according to type of milk. Pearson χ^2 test was employed to analyze qualitative categories, whilst rates $p \le 0.05$ were significant.

III. RESULTS AND DISCUSSION

Since the commercialization of raw milk is illegal, Brazilian legislation has not provided parameters for the product. When Norm 62 published on 29 December 2011 (Brasil, 2011) on refrigerated raw milk for processing is applied, it has been found that 80.95% of samples revealed mesophyll microorganisms, of which 76.47% were above the rate allowed by current legislation, or rather, 5.87 log CFU/ mL (Brasil, 2011). Samples from three out of the five towns averaged above the acceptable rate (Table 1).

TABLE 1

MEANS AND VARIATIONS OF MICROBIAL POPULATIONS IN SAMPLES OF RAW MILK INFORMALLY COMMERCIALIZED IN TOWNS OF THE RECÔNCAVO DA BAHIA, BRAZIL, 2015.

Microorganisms (log		Legislation				
CFC/ mL)	A	В	С	D	E	IN 62/2011

Mesophylls	9.17 (7.61 - 9.75)	8.7 (6.6 - 9.75)	6.34 (<1 - 8.75)	5.84 (<1 - 8.75)	5.13 (<1 - 5.13)	5.87
Psichrotrophic	7.11 (<1 - 9.75)	4.93 (4 - 5.48)	4.57 (<1 - 8.75)	3.80 (1.9 - 5.62)	2.18 (<1 - 2.18)	SD
Thermophile	6.17 (<1 - 6.17)	<1	<1	1.62 (<1 - 2.23)	<1	SD
Total coliforms	8.17 (5.86 - 9.75)	6.29 (5.21 - 6.94)	6.56 (4.18 - 8.75)	5.32 (1.6 - 8.75)	5.03 (<1 - 5.03)	SD
E.coli	2.35 (0.95 - 6.2)	1.63 (<1 - 1.95)	3.49 (<1 - 8.75)	1.85 (<1 - 1.85)	1.60 (<1 - 1.60)	SD

Source: Research data; SD: rates without any legal determination

High population of mesophyll microorganisms in the raw milk analyzed may be attributed to inadequate sanitary conditions during milking and mainly to lack of refrigeration in transport, storage and commercialization of the product. It was perceived during collection of samples that most samples lacked refrigeration and directly contributed towards the proliferation of mesophyll microorganisms since best temperature for growth was that of room temperature.

Silva et al. (2010) evaluated the physical, chemical and microbiological quality of raw milk of a dairy farm in the state of Rio Grande do Sul and reported that three out of the six milk samples analyzed were above legal rates (Brasil, 2011) for mesophyll microorganisms. Similarly to current study, Silveira & Bertagnolli (2014) analyzed the quality of raw milk commercialized illegally in the street fairs of Santa MariaRS Brazil and detected that three out of ten samples failed to comply with Norm 51 of 20/09/2002 for mesophyll organisms in refrigerated milk.

On the other hand, samples of UHT milk complied with Rule 146 of 07/03/1996 with rates up to 2 log UFC/ mL for mesophylls. Studies by Frata et al. (2014) provided similar results, or rather; samples were within the maximum mesophyll limit for UHT milk. However, Domareski et al. (2010) assessed UHT milk commercialized in three Mercosur countries (Brazil, Argentina and Paraguay), respectively reported 37.5%, 62.5% and 12.5% mesophyll aerobic bacteria in milk, and thus failing to comply with microbiological criteria for UHT milk.

Storage temperature and type and initial microbial load are parameters that contribute towards the proliferation of bacteria during the storage of raw milk even when submitted to low temperatures. Raw milk kept for long periods at low temperatures may reveal psichrotrophic microorganisms and their concentration in milk may be associated to conditions in which the milk was obtained.

Although Brazilian legislation on the subject (Brasil 2001, 2011) does not establish a maximum limit for psichrotrophic and thermophile microorganisms, the Rules for Industrial and Sanitary Inspection of Animal-derived Products(RIISPOA) (Brasil, 1980) determine that milk should have a maximum of 10% psichrotrophic and thermophile microorganisms with total mesophyll counts

According to the above recommendation, only 9.52% of raw milk samples in current analysis had the best pattern for psichrotrophic microorganisms. Zeni et al. (2013) verified the occurrence of psichrotrophic and mesophyll microorganisms in raw milk in the production of UHT milk. Samples of refrigerated raw milk had mesophyll and psichrotrophic counts above 6 log CFU/mL, which will surely interfere in the quality and shelf life of the final product.

Although raw milk does not normally have high rates of thermophile microorganisms, current study reveals averages between 6.17 log CFU/mL and 1.62 log CFU/mL respectively for municipalities A and D, with 14.28 % of samples above 10% of mesophyll microorganisms, blatantly not complying with current legislation (Brasil, 1980). There was a microbial multiplication of <1 log CFU/mL in all UHY milk samples.

Further, 90.47% of samples had over 5 log UFC/ mL of total coliforms in raw milk. Although there is no maximum limit for total coliforms in raw milk when current sanitary legislation is taken into account, the microbial load is high in the samples. In fact, microorganisms reveal flaws in hygiene control and the possible presence of pathogenic microorganisms.

Corroborating results in current analysis, Dias et al. (2015) assessed the hygiene and sanitary conditions of raw milk commercialized in outdoor markets in the north of the state of Piauí, Brazil, and found that the 16 samples analyzed were positive for total and thermotolerant coliforms.

In a study in Solânea in the state of Paraíba, Brazil, Amaral & Santos (2011) reported that raw milk sold by street vendors had more than 1.100 MPN/mL of total and thermotolerant coliforms. Another study with similar results was performed by Maciel et al. (2008). The authors analyzed 30 samples of raw milk and detected total coliforms in all samples.

Samples from municipalities A and B showed average of 0.87 log CFU/mL and 2.25 log CFU/mL of total coliforms in UHT milk, respectively, and thus not adequate for consumption. In fact, Resolution 370 of 4/9/1997rules that UHT milk should not contain microorganisms which are able to proliferate at normal storage and distribution conditions after the incubation of the closed package at 25-37°C for seven days.

According to Menezes et al. (2015), hygienic milking procedures, cleansing and disinfection of utensils and equipments are basic to avoid milk contamination by coliforms. Training of milk producers with regard to hygiene at all stages in the provision and commercialization of raw milk is mandatory to guarantee the safeness of the product for industrialization.

E. coli was detected in samples of raw milk from all the municipalities and varied between <1 and 1.64 log CFU/ mL, whereas it was detected in UHT milk from one municipality only, averaging 1.46 log CFU/ mL. Its occurrence and its high population in milk are highly relevant for public health since, besides the existence of pathogenic strains, it is the main indicator for feces-caused contamination..

Microbiological analysis of raw milk revealed *Staphylococcus aureus*in 76.19% of all samples analyzed, varying between <1 and 9.0 log CFU/ mL. UHT milk samples had rates lower than <1 log CFU/ mL. Student's t test showed a statistical difference between raw and UHT milk, with higher contamination in the former (p<0.05).

Although there is no maximum limit for *Staphylococcus aureus* in raw milk, its occurrence is associated with lack of hygiene of handlers' hands, utensils used in milking, bad storage, transport and commercialization conditions. The above contamination situation was observed during sampling. In fact, milk could be found in inadequate containers, without any identification, featuring dirt rates. Others lacked refrigeration and this fact may have contributed towards the multiplication of the microorganism in most samples.

One may conclude that average counts for *S.aureus* populations in the milk from the municipalities were similar and showed that the presence and high population rates of *S.aureus* is normal in raw milk (Table 2).

TABLE 2
MEANS AND VARIATIONS OF *STAPHYLOCOCCUS AUREUS* POPULATIONS IN RAW MILK SAMPLES COMMERCIALIZED INFORMALLY IN TOWNS OF THE RECÔNCAVO DA BAHIA, BRAZIL, 2015.

Microorganisms (log CFC/ mL)		Legislation				
	A	В	С	D	E	IN 62/2011
Staphylococcus aureus	6.06 (3.54 - 9.75)	2.70 (1.78 - 4.18)	2.69 (<1 - 4.13)	2.04 (<1 - 2.04)	3.12 (<1 - 3.12)	SD

Source: Research data; SD: rates without any legal determination

Research evaluating the microbiological quality of raw milk informally commercialized in Areia PB Brazil showed 80% of samples were contaminated by *Staphylococcus* sp (30% were presumably *S. aureus*) corroborating results in current study

(Souza et al., 2011). Alves et al. (2009) assessed the microbiological quality of raw milk commercialized in São Luís MA Brazil, and detected strains of *Staphylococcus* positive coagulase in 31.0 % of samples.

Similar results were reported for UHT milk by Nascentes &Araújo (2012) who compared the microbiological quality of raw, pasteurized and UHT milk commercialized in Patos de Minas MG Brazil. The authors reported that all samples of raw milk were contaminated by *S.aureus*, whereas no growth of the microorganism was detected in UHT milk sample.

E.coli O157:H7 occurred in 6.12% and 2.04% of raw and UHT milk samples respectively, or rather, a potential risk for consumers since it is a pathogenic strain. Further, Pearson's χ^2 test showed no significant difference between the types of milk studied with regard to *E.coli* O157:H7 (p>0.05). Since the microorganism is highly susceptible to heat, it may be easily destroyed during milk processing. However, post-processing contamination may occur due to its ability to form biofilms on utensils and equipments. According to Pillai & Jesudhasan (2006), the ability of *E.coli* O157:H7 in forming biofilms may be attributed to the self-induction-2 signal involving the regulation of genes in chemotaxis, flagellar synthesis and motility.

Batista et al. (2014) evaluated the hygiene and sanitary parameters of raw milk on 26 farms in the region of the Recôncavo da Bahia BA Brazil and reported one positive sample of *E.coli* O157:H7.

Above data show the importance of permanent studies on raw and processed milk, underscoring the need for the non-commercialization of raw milk directly to the consumer and the hygiene conditions during the post-processing stage.

It should be emphasized that *E. coli* O157:H7 in UHT milk is a serious factor due to the fact that milk had undergone commercial sterilization process and the consumer is liable to be contaminated by the pathogenic microorganism by purchasing an unsafe product.

In the case of *Listeria monocytogenes*, the result in all milk samples proved to be negative. In a study on *Listeria* spp. in raw and pasteurized milk, 10% of raw milk samples were contaminated by the microorganism, with no contamination of any pasteurized milk sample (Almeida et al., 2013).

Although current analysis did not reveal *Listeria monocytogenes*, its presence in milk is of great concern due to the microorganism's high pathogenicity which causes listeriosis worldwide. Lack of detection of the pathogenic microorganism may also be related to its low capacity of competition by nutrients and thus undetectable, albeit present.

When all the aspects evaluated and results for microorganisms which indicate sanitary and pathogenic quality in raw milk in the municipalities under analysis are taken into account, the samples do not comply with current Brazilian legislation for refrigerated raw milk (Brasil, 2011) with regard to microbiological aspects. In fact, it is risk for public health and may be a FTD vector.

Good Practices in handling should be practiced by producers to reduce diseases caused by the incorrect handling of raw milk on the farms and in storage, transport and commercialization of the final product after a correct processing. Efficient measures for the eradication of the sale of raw milk should be aimed at by the health authorities since no commercialization of such a product should occur.

When the microbiological quality of UHT milk samples commercialized in the municipalities evaluated is assessed, several samples were not fit for consumption, possibly by post-processing microbial contamination by pathogens *E.coli* and *E.coli* O157:H7.

Since the microorganisms analyzed were not resistant to extreme temperatures such as in UHT milk processing, it may be suggested that samples with pathogenic microorganisms were contaminated after thermal processing either during packaging or due to biofilms on the equipments or deficient processing. According to Vittori et al. (2008), post-processing contamination may be due to deficiency in packaging sterilization or to recontamination by handlers. The emergence of biofilms in milk processing environments contributes towards an increase in the probability of microbial contamination of processed milk products with deteriorating and pathogenic microorganisms (Marchand et al., 2012).

Bad quality of raw milk used as prime matter and preservation problems during commercialization may also cause microbial contamination in processed milk with the possibility of milk deterioration prior to the recommended preservation period (Dey & Karim, 2013).

Dairy industries should guarantee the quality of the product by sanitary control during the whole processing period, from the acquisition of prime matter to sale.

IV. CONCLUSION

- The commercialization of raw milk is highly dangerous for consumers due to rates of pathogenic microorganisms found in it.
- Contamination of UHT milk may have been due to flaws in post-processing sanitary control that favored strains of total coliforms, *E.coli* and *E.coli* O157:H7 in the commercialized milk.
- Negative results occurred for *L. monocytogenes* in all samples under analysis.

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