Interspecific Variation in Salt Tolerance of Some Acacia Species at Seed Germination Stage

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Abstract— Germination of seeds from six Acacia species was evaluated under salt stresses using five treatment levels: 0, 100, 200, 300, and 400mM of NaCl. Corrected germination rate (GC), germination rate index (GRI) and mean germination time (MGT) were recorded during 10 days. The results indicate that germination was significantly reduced in all species with the increase in NaCl concentrations. However, significant interspecific variation for salt tolerance was observed. The greatest variability in tolerance was observed at moderate salt stress (200 mM of NaCl) and the decrease in germination seems to be more accentuated in A. cyanophylla and A. cyclops. Although, A. raddiana, remains the most interesting, it preserved the highest percentage (GC = 80%) and velocity of germination in all species studied in this work, even in the high salt levels. This species exhibits a particular adaptability to salt environment, at least at this stage in the life cycle, and could be recommended for plantation establishment in salt affected areas. On the other hand, when ungerminated seeds were transferred from NaCl treatments to distilled water, they recovered largely their germination without a lag period, and with high speed. This indicated that the germination inhibition was related to a reversible osmotic stress that induced dormancy rather than specific ion toxicity.

Keywords— Acacia species, Osmotic stress, Germination recovery, Salt tolerance, Seed germination, Variability, Plant breeding, Rehabilitation, Salt areas.

I. INTRODUCTION

Salinity of soils is one of the most environmental factors limiting agricultural production and has significant effects on crop productivity and biodiversity. It has more severe impact in arid and semi-arid environments and combined with the water constraint presents a serious threat to food stability in these areas [1,2,3]. Indeed, salinization already has affected more than 800 million hectares throughout the world or 6% of total land [4,5]. In the North Africa and the Middle East, salinity affects 15 million hectares of arable lands and this area is in continuous progression [6,7]. In Morocco, more than 5% of areas are already affected by salinity to various degrees [8]. They correspond especially to arid and semi-arid regions where 80% of available water for irrigation contains salinity higher or equal to 5 g/l [9]. These wide geographical areas are not exploited to a great extent, except when they occasionally constitute a poor pasture land or irrigated domain with low yield.

The best economic approach for exploitation and rehabilitation of these marginal regions is selection of salt tolerant species and varieties capable of sustaining a reasonable yield within salt-affected soils [10,11,12]. The effectiveness of such approach depends on the availability of genetic variation in relation with salt tolerance and its exploitation by screening and selection of the powerful plants under saline stress [13,14,15]. For many crop species, intraspecific variability for salt tolerance has been identified among cultivars and wild species [16,17,18,19]. These rustic resources are well adapted and constitute a potential reservoir useful to provide interesting materials in order to diversify and increase the productivity, particularly in pasture land affected by salinity.

Acacia species have the ability to survive in a diverse range of habitat and environments. They are well adapted to the arid and semi-arid regions and are known for them tolerance to high pH and salinity as well [20,21]. These species are able to stabilize and fertilize soils via nitrogen fixing and mycorrhizal symbioses [22,23] and constitute sources of wood, fodder, medicine and gum [24]. Acacia trees may thus include ideal candidates for enabling saline land reclamation with a potential for financial returns because of their combined production and soil improvement [25,26,27].

In Morocco, the genus *Acacia* is represented by four spontaneous species including one Moroccan endemic (*Acacia* gummifera Wild.); the other three are *Acacia raddiana* Savi, *Acacia ehrenbergiana* Hayne., *Acacia albida* Del. Otherwise, about ten species are introduced for ornamentation, reforestation or the fight against desertification [28].

Successful establishment of plants often depends on germination success, especially those that grow in salt affected areas. Thus, seeds must remain viable for long period in high salinity conditions and germinate when salinity decreases [29]. Various halophytic or glycophytic species, show an important variability with their ability to germinate under different salt concentration [30,31,32]. The effects on germination depend on the concentration of NaCl and varied among the plant species [33]. To overcome salt stress effect, plants have evolved various mechanisms that help them to adapt the osmotic and ionic stress caused by high salinity [26]. Otherwise, salts can affect germination of seeds either by restricting the supply of water (osmotic effect) or by causing specific injury through ions to the metabolic machinery (ionic effect) [34].

The purpose of this study was to assess and compare the seed germination response of six *Acacia* species (*Acacia gummifera*, *Acacia raddiana*, *Acacia eburnea*, *Acacia cyanophylla*, *Acacia cyclops* and *Acacia horrida*) under different NaCl concentrations in order to explore opportunities for selection and breeding salt tolerant genotypes that can be utilized in future land reclamation projects. The study will also assess to determine whether salt stress is induced by osmotic constraints or by toxic effect of NaCl.

II. MATERIALS AND METHODS

2.1 Plant Material

Six Acacia species were studied: Tow spontaneous species (Acacia gummifera and Acacia raddiana) and four introduced species (Acacia eburnea, Acacia cyanophylla, Acacia cyclops and Acacia horrida). Mature, dry seeds were collected from trees growing under field conditions in semi-arid region of south Moroccan region. Seeds collected from pods were generously provided by the Regional Forest Seeds Station of Marrakech and stored in a cold chamber at 6°C.

Acacia gummifera is an endemic species to Morocco and Acacia tortilis (Forssk.) Hayne subsp. raddiana (Savi) Brenan, commonly named A. raddiana, considered as a keystone species, is more prevalent in the inland part of the ecoregion, and are widely distributed throughout the Sahara desert [35,36]. The other four species were introduced in Morocco from southwestern Australian in the 18th century. These plantations were created for several purposes such as their use as ornamental plant, in the fight against desertification and for dune stabilization [37].

2.2 Germination

Seeds from different pods were manually scarified, to overcome hard seed coat dormancy, and sterilized with 0.5% sodium hypochlorite solution (NaOCl) for 10 minutes, then rinsed with sterile distilled water several times, and briefly blotted on filter paper. Three replicates of 20 seeds from each accession were placed in plastic Petri dishes (90 mm diameter) on filter paper wetted with distilled water (control) and four salinity concentrations (100, 200, 300 and 400 mM NaCl). Petri dishes were randomized in a precision incubator and maintained in the dark at 25 ± 0.5 °C. Seeds were considered to have germinated when their radicle reached at least 3 mm long. Germination response was recorded daily for 10 days.

Several germination parameters were calculated to characterize the salt tolerance, including the corrected germination rate (GC), germination rate index (GRI) and mean germination time (MGT):

- Corrected germination rate (GC) was expressed as the number of seeds germinated in a concentration of salt divided by the number of germinated seeds in distilled water (control) for 10 days [38].
- The germination rate index (GRI) was calculated by using the following formula:

$$GRI = (G1 / 1) + (G2 / 2) + ... + (Gx / x)$$

Where G is the germination percentage at each day after sowing, and 1, 2, ..., x represents the corresponding day of germination. The value of GRI was higher when seeds germinated earlier. This parameter described by [39,40] is a measure of seedling vigor and should involve not only germination but emergence characteristics.

• The Mean Germination Time (MGT) is a measure of the rate and time-spread of germination (lower values indicating faster germination). It was estimated as:

$$MGT = \sum ni-ti/N$$

where t is time from the beginning of the germination test in terms of days and n is the number of newly germinated seeds at time t [41,42].

To test germination recovery performance after salt exposure, ungerminated seeds in severe salt stress (300 and 400 mM of NaCl) were transferred to distilled water and incubated for 6 days. We calculated the recovery germination percentage (number of germinated seeds in recovery test / number of seeds transferred to distilled water) [43].

2.3 Statistical analysis

All values expressed as a percentage were arcsine square root transformed before performing statistical analysis to normalize the data and improve homogeneity of variance [44]. These measures were submitted to a two ways analysis of variance (ANOVA) with species and salinity treatments as factors followed by a Student–Newman–Keuls post hoc test. A difference was considered to be statistically significant when P < 0.05. All statistical analysis were performed with Statistica software Version 6.1 for Windows [45].

III. RESULTS AND DISCUSSION

3.1 Effect of salinity on seed germination

For the six species, the two-way ANOVA indicated highly significant main effect of both species and salinity regarding final germination percentage and germination rate index (P<0.001) (Table 1 and 2). However the existence of a significant interaction between these two effects (F = 34, 00 at (GC) and F = 6,385 at (GRI)) indicates that species studied did not similarly respond to the effect of salt at a given concentration of NaCl.

TABLE 1
TWO-WAY VARIANCE ANALYSIS (ANOVA) FOR FINAL GERMINATION PERCENTAGE (GC) OF THE SIX SPECIES UNDER DIFFERENT CONCENTRATIONS OF NaCl.

Source of variation	d. f.	MC	F
Species (Sp.)	5	0,618	270,110**
Salinity (Salt.)	4	5,113	2233,421**
Sp. X Salt	20	0,077	34,000**
Error	60	0,002	

d.f.: Degree of freedom, MC: Mean square, F: ratio of variances, **significant at 1% probability level.

TABLE 2
TWO-WAY VARIANCE ANALYSIS (ANOVA) FOR GERMINATION RATE INDEX (GRI) OF THE SIX SPECIES UNDER DIFFERENT CONCENTRATIONS OF NaCl.

Source of variation	d. f.	MC	F	
Species (Sp.)	5	0,570	283,194**	
Salinity (Salt.)	4	4,149	2064,370**	
Sp. X Salt	20	0,040	20,010**	
Error	60	0,002		

d.f.: degree of freedom, MC: Mean Square, F: ratio of variances, **significant at 1% probability level.

For each concentration of NaCl, a significant interspecific variation in both potential of seed germination (fig. 1) and seedling vigor (fig. 2) among the six examined species. Mean comparison at different salinity levels indicated that increase of salinity causes a decrease in seed germination capacity, which was higher in distilled water than in any NaCl concentration. Moreover, all the species showed an increase in MGT (Mean Germination Time), indicating that seeds germinated more slowly as salinity increased (Table 3).

TABLE 3
MEAN GERMINATION TIME (MGT) OF DIFFERENT SPECIES IN VARIOUS NaCl CONCENTRATIONS.

Species	NaCl Concentrations					
	0mM	100mM	200mM	300mM	400mM	
A. cyanophylla	2,48 a	2,98 a	5,83 b	NC	NC	
A. cyclops	2,58 a	3,09 a	5,42 b	6,67 c	NC	
A. eburnea	3,20 a	3,80 a	4,31 a	5,55 b	7,00 c	
A. gummifera	3,05 a	3,48 a	4,24 a	5,66 c	6,22 c	
A. horrida	3,03 a	3,58 ab	4,76 b	6,50 c	NC	
A. raddiana	2,66 a	2,80 a	3,30 a	4,86 b	6,08 c	

Values followed by the same letter in a row are not significantly different at P<0.05. (Newman-Keuls test). NC: MGT was not calculated because of insufficient germination

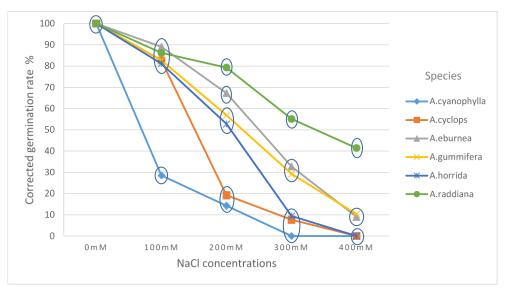


FIG. 1: CORRECTED GERMINATION RATE OF DIFFERENT SPECIES IN VARIOUS NaCl CONCENTRATIONS. At each concentration of NaCl, values in the same ellipse are not significantly different (p < 0.05) (Newman-Keuls test).

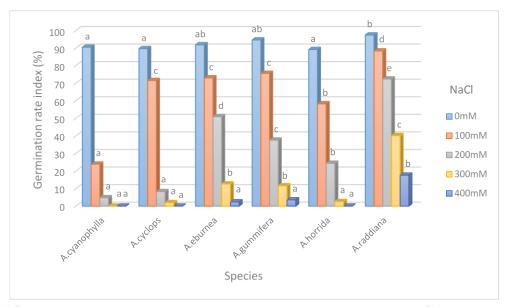


FIG. 2: GERMINATION RATE INDEX OF DIFFERENT SPECIES IN VARIOUS NaCl CONCENTRATIONS. At each concentration of NaCl, means of populations having the same letter are not significantly different (p < 0.05) (Newman-Keuls test).

At the lowest stress (100 mM), the decrease in germination seems to be more accentuated in *A. cyanophylla* (GC never fell above 30%). However, all the other species are distinguished by forming a homogeneous group which is little affected by salinity compared with control (GC = 84% on average). At this saline level, seeds germinated rapidly and no significant change was noticed in germination speed (MGT not differing from that of control) (Table 3).

At 200 mM, the results revealed considerable interspecific variation in the response of seed germination to salinity among the studied species. Thus, four groups are distinguished, the first is formed by *A. raddiana* which showed very high germination percentage (GC = 80%) and revealed earlier seed germination MGT = 3,29, followed by *A. eburnea* (GC and GRI close respectively to 67% and 51%), while *A. gummifera*, and *A. horrida* occupy an intermediate position between the tolerant species and the rest of the others, considered as the most sensitive (GC = 15% in the case of *A. cyanophylla* and 19% for *A. cyclops*). Otherwise, from this level, seed germination was relatively slower in *A. cyanophylla* and *A. cyclops* (high MGT).

The elevated doses of salt (300 and 400 mM) induced significant reduction in seed germination and retarded their initiation of all the species. A. cyanophylla, A. horrida, and A. cyclops seem to be sharply affected in the same manner by these two

concentrations, their corrected germination percentage does not exceed 6% and the time to germination gradually lengthened (MGT> 6 days) at 300 mM and no germination took place in this group at 400 mM. Seeds of *A. gummifera* and *A. eburnea* can be regarded as moderately tolerant to salt stress and react in the same way at these two concentrations (GC and GRI close respectively to 32% and 12% in the case of 300 mM). In this case germination was significantly delayed at the tow highest NaCl levels.

Whereas, *A. raddiana* continued to record highest percentage and velocity of germination in all species studied in this work, even in the high salt levels with GC reached 55% at 300 mM and 41% at 400 mM of NaCl. This species exhibits a particular adaptability to salt environment, at least at this stage in the life cycle, and could be recommended for plantation establishment in salt affected areas.

The effect of salinity on germination has been addressed by several authors and in different species [46,47,34,48,49]. This stage is very important for the development of the plants, particularly those that live in environments affected by salinity. In this study, the monitoring of germination process revealed that salinity notably affected germination in *Acacia* species but also delayed the time needed to complete germination, especially with increasing salinity level. This is consistent with our result particularly for sensitive species and is explained by the time required for the seeds to develop mechanisms allowing to adjust them internal osmotic pressure [50].

Additionally, the results showed significant interspecific variation in salt tolerance during germination of the species studied in the range of concentrations of sodium chloride from 100 to 400 mM. This variability, required to start a breeding programs for salt tolerance, have been also observed in several species including halophyte or glycophyte species [51,52,43,15,49].

The Moroccan *Acacia raddiana* particularly, tolerate salinity until 400 mM (probably also at higher levels) with a germination rate that exceeds 40%. Indeed, according to Ndour and Danthu [53] and Danthu and al. [54], *Acacia raddiana* is among the African *Acacia* species whose germination is less affected by the presence of salt. Its seed germination could be blocked only up to salt concentrations close to seawater (35 g/l). Previous studies have also reported that this species were the most tolerant and could be used in increasing forage production in salt affected areas [55,56,57,58]. Furthermore, the observed germination percentages in our study are relatively well higher than those published by Jaouadi and al. [50] in Tunisian species (maximum germination rate of 21% under 22 g/l of NaCl) and by Abari and al. [59] in Iranian taxa (germination was stopped at 300 mM). El Nour and al. [46] reported that no germination took place in four Yemeni *Acacias* (*A. cyanophylla*, *A. seyal*, *A. tortilis*, and *A. tumida*) at salt concentration above 20 dSm⁻¹ (around 200 mM of NaCl).

Among the rest of the five *Acacia* species examined in the present study, *A. gummifera* and *A. eburnea* was shown to possess the medium level of salt tolerance, whereas *A. cyanophylla*, *A. horrida*, and *A. cyclops* were categorized as the least tolerant group. Several papers that have compared salt tolerance in some *Acacia* species have suggested relatively the same ranking [60].

Moreover, previous studies have reported that salt tolerance was positively correlated with seed size [61,26]. *A. raddiana*, *A. gummifera* and *A. eburnea* seeds are larger than those of *A. cyanophylla*, *A. horrida*, and *A. cyclops*; and according to Croser and al. [62], larger seeds may contain more food reserves, which could be used to overcome osmotic effects of salts, and greater energy reserves making them less dependent on photosynthesis for early growth. Similar observation which made on *Triticum aestivum* [63,64], on *Atriplex* species [65] as well as on *Acacia longifolia* [26], which also showed that larger seeds had greater success in overcoming osmotic constraints during the initial stages of germination.

El Nour and al. [46] reported that salinity notably delayed the germination process of *Acacia* species, especially with increasing salinity level. This is consistent with our result particularly for sensitive species and explained by the time required for the seeds to develop mechanisms allowing to adjust them internal osmotic pressure [50].

3.2 Recovery of Germination

After transfer of the seeds that failed to germinate under high concentrations of salt to distilled water, they recovered relatively their aptitude of germination at all the species studied. The delay in germination speed tended to be relatively rapid than that observed in distilled water (fig. 3).

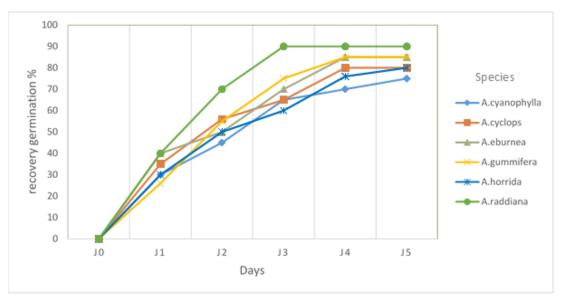


FIG. 3. THE REVERSIBLE EFFECT OF HIGH NaCl CONCENTRATION ON THE GERMINATION KINETICS IN THE STUDIED SPECIES.

This revealed that NaCl had no toxic effect, because salt stress did not damage the embryo as verified by not only the germination recovery but also pink embryos in the tetrazolium staining test. NaCl concentrations did not destroy seed germination ability, it had only repressed the germination momentarily and the viability was preserved. These results are consistent with those obtained in several other salt tolerant species including *Atriplex halimus* [38], *Medicago ruthenica* [66], *Medicago polymorpha* and *Medicago ciliaris* [67], *Panicum turgidum* [30], *Triticum aestivum* [68], *Brassica napus* [19] and *Lolium perenne* and *Bromus tomentellus* [69].

An important characteristic of salt tolerant seeds, which differentiate them from seeds of glycophytes, is their aptitude to maintain seed viability for lengthy period of time during exposure to hyper-saline conditions and then initiate germination when salinity stress is decreased [70,71,30]. The present study reveals that the recovery of germination is not a criterion of salt tolerance which distinguishes halophytes from glycophytes. It was maybe due to a reversible osmotic effect that induced dormancy, as revealed in findings of [72,68]. Consequently, a high proportion of seeds remained viable and had the ability to germinate when salinity stress was alleviated [30].

Reduction in mechanisms of germination by osmotic stress may be related to the lower diffusion of water through the seed coat [73,47] caused by the increased osmotic pressure environment, preventing the seed imbibition [65]and mobilization of reserves for embryo's growth [74]. Thus, dormancy decrease the risk of seedling mortality when moisture is limited and salinity is augmented [75]. High recovery germination speed revealed in our studied species, particularly in *A. raddiana*, indicates that seeds have the ability to avoid deterioration caused by prolonged exposure to unfavorable biotic factors [76]. This situation constitutes an ecophysiological adaptive strategy to take full advantages of favorable conditions, available for a short time, during the germination stage [77]. It also, secure the long-term subsistence of seed bank helping the species in dispersal germination and seedling establishment over years [75]. Tilaki and al. [69] revealed that under saline environments, seed survival may be a suitable condition for success instead of germination capacity, since recovery germination does occur in the seeds when hyper-saline conditions are alleviated.

IV. CONCLUSION

The present study, indicate that salinity decreased rate and speed of germination. It reveals also significant interspecific variation in the potential for salt tolerance at germination among the six species. The Moroccan species *Acacia raddiana* remains the most interesting at this stage. It preserves a high rate of germination until a concentration 400 mM of NaCl, with a germination rate that exceeds 40%. Germination recovery after removing the saline stress indicates that sodium chloride caused a reversible osmotic effect of germination rather than ion specific toxicity and exerts a temporary inhibition of germination which is eliminated with the removal of the constraint. This ability to germinate after exposure to higher concentrations of NaCl suggest that these species, especially the most tolerant, could be able to germinate under the salt affected soils and could be utilized for the rehabilitation of damaged arid zones. Nevertheless, germination in field conditions

is more difficult due to other environmental factors such as, drought, light, and temperature. A more ambitious program, including *Acacia* species, is necessary, not only at germination but also at the other stages of the life cycle. This opens the possibility to continue this work to verify correlation between salt tolerance during seed germination and early stage of plant development which will be most useful in a breeding program for selecting salt-tolerant genotypes in *Acacia* species.

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REFERENCES

- [1] Ashraf, M. and M. Foolad, 2007, Roles of glycine betaine and proline in improving plant abiotic stress resistance. Environmental and Experimental Botany, 59(2), 206-216
- [2] Porcel, R., R. Aroca, R. Azcon and J.M. Ruiz-Lozano, 2016, Regulation of cation transporter genes by the arbuscular mycorrhizal symbiosis in rice plants subjected to salinity suggests improved salt tolerance due to reduced Na+ root-to-shoot distribution. Mycorrhiza, 1-12
- [3] Rojas, R.V., M. Achouri, J. Maroulis and L. Caon, 2016, Healthy soils: a prerequisite for sustainable food security. Environmental Earth Sciences, 75(3), 1-10
- [4] Cramer, G.R., K. Urano, S. Delrot, M. Pezzotti and K. Shinozaki, 2011, Effects of abiotic stress on plants: a systems biology perspective. BMC plant biology, 11(1), 163
- [5] Wu, H., L. Shabala, X. Liu, E. Azzarello, M. Zhou, C. Pandolfi, Z.-H. Chen, J. Bose, S. Mancuso and S. Shabala, 2015, Linking salinity stress tolerance with tissue-specific Na(+) sequestration in wheat roots. Frontiers in Plant Science, 6, 71
- [6] Lahmar, R. and A. Ruellan, 2007, Dégradation des sols et stratégies coopératives en Méditerranée: la pression sur les ressources naturelles et les stratégies de développement durable. Cahiers Agricultures, 16(4), 318-323
- [7] Bui, E., 2013, Soil salinity: A neglected factor in plant ecology and biogeography. Journal of arid environments, 92, 14-25
- [8] Barbouchi, M., R. Abdelfattah, K. Chokmani, N. Ben Aissa, R. Lhissou and A. El Harti, 2015, Soil Salinity Characterization Using Polarimetric InSAR Coherence: Case Studies in Tunisia and Morocco. Journal of Selected Topics in Applied Earth Observations and Remote Sensing, IEEE, 8(8), 3823-3832
- [9] Choukr-Allah, R., 1991. The use of halophytes for the agricultural development of south of Morocco. Proc Int Conf Agric Manag salt affected areas, Agadir. 25,377-386
- [10] Ghoulam, C. and K. Fares, 2001, Effect of salinity on seed germination and early seedling growth of sugar beet (Beta vulgaris L.). Seed science and Technology, 29(2), 357-364
- [11] Ashraf, M.Y., A.R. Awan and K. Mahmood, 2012, Rehabilitation of saline ecosystems through cultivation of salt tolerant plants. Pak. J. Bot, 44, 69-75
- [12] Mirzabaev, A., M. Ahmed, J. Werner, J. Pender and M. Louhaichi, 2016, Rangelands of Central Asia: challenges and opportunities. Journal of Arid Land, 8(1), 93-108
- [13] Singh, S., R. Sengar, S. Bhatnagar, P. Chand, M. Yadav and R. Singh, 2014, Assessment of genetic variability of bread wheat (Triticum aestivum L.) genotypes for salinity using salt tolerance indices. International Journal of Agriculture Innovations and Research, 3(3), 874-879
- [14] Chattopadhyay, K., D. Nath, R. Mohanta, B. Marndi, D. Singh and O. Singh, 2015, Morpho-physiological and Molecular Variability in Salt Tolerant and Susceptible Popular Cultivars and their Derivatives at Seedling Stage and Potential Parental Combinations in Breeding for Salt Tolerance in Rice. Cereal Research Communications, 43(2), 236-248
- [15] Joseph, S., D.J. Murphy and M. Bhave, 2015, Identification of salt tolerant Acacia species for saline land utilisation. Biologia, 70(2), 174-182
- [16] Correia, P., F. Gama, M. Pestana and M. Martins-Loução, 2010, Tolerance of young (Ceratonia siliqua L.) carob rootstock to NaCl. Agricultural water management, 97(6), 910-916
- [17] Daffalla, H.M., R.S. Habeballa, E.A. Elhadi and M.M. Khalafalla, 2013, Random amplified polymorphic DNA (RAPD) marker associated with salt tolerance during seeds germination and growth of selected Acacia senegal provenances. African Journal of Biotechnology, 10(31), 5820-5830
- [18] Tiwari, R.S., G.A. Picchioni, R.L. Steiner, D.C. Jones, S. Hughs and J. Zhang, 2013, Genetic variation in salt tolerance at the seedling stage in an interspecific backcross inbred line population of cultivated tetraploid cotton. Euphytica, 194(1), 1-11
- [19] Yong, H.-Y., C. Wang, I. Bancroft, F. Li, X. Wu, H. Kitashiba and T. Nishio, 2015, Identification of a gene controlling variation in the salt tolerance of rapeseed (Brassica napus L.). Planta, 242(1), 313-326
- [20] Abbas, G., M. Saqib, J. Akhtar and S.M.A. Basra, 2013, Salinity tolerance potential of two Acacia species at early seedling stage. Pakistan Journal of Agricultural Sciences, 50 (4), 683-688
- [21] Bui, E.N., A. Thornhill and J.T. Miller, 2014, Salt-and alkaline-tolerance are linked in Acacia. Biology letters, 10(7), 20140278
- [22] Duponnois, R., C. Plenchette, Y. Prin, M. Ducousso, M. Kisa, A.M. Ba and A. Galiana, 2007, Use of mycorrhizal inoculation to improve reafforestation process with Australian Acacia in Sahelian ecozones. Ecological Engineering, 29(1), 105-112

- [23] Sakrouhi, I., M. Belfquih, L. Sbabou, P. Moulin, G. Bena, A. Filali-Maltouf and A. Le Quéré, 2016, Recovery of symbiotic nitrogen fixing acacia rhizobia from Merzouga Desert sand dunes in South East Morocco–Identification of a probable new species of Ensifer adapted to stressed environments. Systematic and applied microbiology, 39(2), 122-131
- [24] Sakrouhi, I., M. Belfquih, L. Sbabou, P. Moulin, G. Bena, A. Filali-Maltouf and A. Le Quéré, 2016, Recovery of symbiotic nitrogen fixing acacia rhizobia from Merzouga Desert sand dunes in South East Morocco – Identification of a probable new species of Ensifer adapted to stressed environments. Systematic and Applied Microbiology, 39(2), 122-131
- [25] Yokota, S., 2003, Relationship between salt tolerance and proline accumulation in Australian acacia species. Journal of Forest Research, 8, 89-93
- [26] Morais, M.C., M.R. Panuccio, A. Muscolo and H. Freitas, 2012, Does salt stress increase the ability of the exotic legume Acacia longifolia to compete with native legumes in sand dune ecosystems? Environmental and Experimental Botany, 82, 74-79
- [27] El Tahir, B.A., M. Daldoum and J. Ardö, 2013, Nutrient Balances as Indicators of Sustainability in acacia senegal Land use Systems in the Semi-arid Zone of North Kordofan, Sudan. Standard Scientific Research and Essays, 1(5), 93-112
- [28] Lahdachi, F.Z., L. Nassiri, J. Ibijbijen and F. Mokhtari, 2015, Aperçu sur les acacias spontanés et introduits au Maroc. European Scientific Journal, 11(23), 88-102
- [29] Tobe, K., X. Li and K. Omasa, 2000, Seed germination and radicle growth of a halophyte, Kalidium caspicum (Chenopodiaceae). Annals of Botany, 85(3), 391-396
- [30] El-Keblawy, A., 2004, Salinity effects on seed germination of the common desert range grass, Panicum turgidum. Seed Science and Technology, 32(3), 873-878
- [31] Kaya, D.M., S. Bayramin, G. Kaya and O. Uzun, 2011, Seed vigor and ion toxicity in safflower (Carthamus tinctorius L.) seedlings produced by various seed sizes under NaCl stress. Archives of Biological Sciences, 63(3), 723-729
- [32] Ruiz, K.B., S. Biondi, E.A. Martínez, F. Orsini, F. Antognoni and S.-E. Jacobsen, 2015, Quinoa-a Model Crop for Understanding Salt-tolerance Mechanisms in Halophytes. Plant Biosystems-An International Journal Dealing with all Aspects of Plant Biology, 150(2), 357-371
- [33] Estrelles, E., E. Biondi, M. Galiè, F. Mainardi, A. Hurtado and P. Soriano, 2015, Aridity level, rainfall pattern and soil features as key factors in germination strategies in salt-affected plant communities. Journal of Arid Environments, 117, 1-9
- [34] Shahriari, A., 2012, Salt tolerance at germination of two forage grasses for reclamation of salinity habitats. Modern Applied Science, 6(7), 36-42
- [35] Munzbergova, Z. and D. Ward, 2002, Acacia trees as keystone species in Negev desert ecosystems. Journal of Vegetation Science, 13(2), 227-236
- [36] Blanco, J., D. Genin and S.M. Carrière, 2015, The influence of Saharan agro-pastoralism on the structure and dynamics of acacia stands. Agriculture, Ecosystems & Environment, 213, 21-31
- [37] Jelassi, A., A. Zardi-Bergaoui, A. Ben Nejma, M. Belaiba, J. Bouajila and H. Ben Jannet, 2014, Two new unusual monoterpene acid glycosides from Acacia cyclops with potential cytotoxic activity. Bioorganic & Medicinal Chemistry Letters, 24, 3777-3781
- [38] Abbad, A., A. El Hadrami and A. Benchaabane, 2004, Germination responses of the Mediterranean saltbush (Atriplex halimus L.) to NaCl treatment. J. Agron, 3(2), 111-114
- [39] Weng, J.-H. and F.-H. Hsu, 2006, Variation of germination response to temperature in formosan lily (Lilium formosanum Wall.) collected from different latitudes and elevations in Taiwan. Plant production science, 9(3), 281-286
- [40] Mirzamasoumzadeh, B., V. Mollasadeghi and S. Elyasi, 2015, Investigating the Effect of Salinity and Drought Stress on Germination of Different Sugar Beet Cultivars Using Correlation between Characters. Research Journal Of Fisheries And Hydrobiology, 10(11), 15-19
- [41] Heydari, M., H. Pourbabaei, O. Esmaelzade, D. Pothier and A. Salehi, 2013, Germination characteristics and diversity of soil seed banks and above-ground vegetation in disturbed and undisturbed oak forests. Forest Science and Practice, 15(4), 286-301
- [42] Shahriari, A. and A. Davari, 2015, The Effect of drought and salinity stresses on seed germination of Alyssum hamalocarpum in Iran's arid lands. International Journal of Agricultural Technology, 11(7), 1625-1639
- [43] Zhang, H., G. Zhang, X. Lü, D. Zhou and X. Han, 2014, Salt tolerance during seed germination and early seedling stages of 12 halophytes. Plant and Soil, 388(1), 229-241
- [44] Turkington, R., E. John, S. Watson and P. Seccombe- Hett, 2002, The effects of fertilization and herbivory on the herbaceous vegetation of the boreal forest in north- western Canada: a 10- year study. Journal of Ecology, 90(2), 325-337
- [45] StatSoft, I., 2001, STATISTICA (data analysis software system), version 6. Tulsa, USA, 150
- [46] El Nour, M., A.A. Khalil and E. Abdelmajid, 2006, Effect of Salinity on Seed Germination Characteristics of Five Arid Zone Tree Specie. U. of KJ Agric. Sci, 14, 23-31
- [47] Bahrami, H. and J. Razmjoo, 2012, Effect of salinity stress (NaCl) on germination and early seedling growth of ten sesame cultivars (Sesamum indicum L.). International Journal of AgriScience, 2(6), 529-537
- [48] Adam Ali Ahmed, N., 2015. Effect of Different Dilution of Red Sea Water on Germination and performance of Seedling of Some Acacia Tree Species. M.Sc in (Desertification), Khartoum University Khartoum (Sudan), 69 p
- [49] Shila, A., M.A. Haque, R. Ahmed and M. Howlader, 2016, Effect of different levels of salinity on germination and early seedling growth of sunflower. World, 3(1), 048-053
- [50] Jaouadi, W., L. Hamrouni, N. Souayeh and M.L. Khouja, 2010, Study of Acacia tortilis seed germination under different abiotic constraints. Biotechnology, Agronomy, Society and Environment, 14, 643-652

- [51] Jaouadi, W., K. Mechergui, L. Hamrouni, M. Hanana and M.L. Khouja, 2013, Effet des contraintes hydrique et saline sur la germination de trois espèces d'Acacias en Tunisie. Revue d'écologie, 68, 133-141
- [52] Yu-ping, R., C. Zhe, Z. Ling-ling, Z. Min, Z. Yong-juan and B. Ke-yu, 2014, Genetic variation patterns of Medicago ruthenica populations from Northern China. African Journal of Biotechnology, 11, 6011-6017
- [53] Ndour, P. and P. Danthu, 1998. Effet des contraintes hydrique et saline sur la germination de quelques Acacias africains. L'acacia au Sénégal. C. Campa, C. Grignon, M. Gueye, S. Hamon and D. S. L'Acacia au Sénégal: Réunion Thématique, 1996/12/03-05. Paris, ORSTOM: 105-122
- [54] Danthu, P., J. Roussel and M. Neffati, 2003. La graine et la germination d'Acacia raddiana. Un arbre au désert : Acacia raddiana. M. Grouzis and E. Le Floc'h. Paris, IRD: 265-283
- [55] Rehman, S., P. Harris, W. Bourne and J. Wilkin, 1997, The effect of sodium chloride on germination and the potassium and calcium contents of Acacia seeds. Seed Science and Technology, 25(1), 45-57
- [56] Rehman, S., P. Harris and W. Bourne, 1998, Effects of presowing treatment with calcium salts, potassium salts, or water on germination and salt tolerance of Acacia seeds. Journal of plant Nutrition, 21(2), 277-285
- [57] Essendoubi, M., F. Brhada, J.E. Eljamali, A. Filali- Maltouf, S. Bonnassie, S. Georgeault, C. Blanco and M. Jebbar, 2007, Osmoadaptative responses in the rhizobia nodulating Acacia isolated from south- eastern Moroccan Sahara. Environmental microbiology, 9(3), 603-611
- [58] Jaouadi, W., K. Mechergui, L. Hamrouni, M. Hanana and M.L. Khouja, 2013, Effet des contraintes hydrique et saline sur la germination de trois espèces d'Acacias en Tunisie. Revue d'écologie, 68(2), 133-141
- [59] Abari, A.K., M.H. Nasr, M. Hojjati and D. Bayat, 2011, Salt effects on seed germination and seedling emergence of two Acacia species. African Journal of Plant Science, 5, 52-56
- [60] Jaouadi, W., L. Hamrouni, M. Hanana and M.L. Khouja, 2004. Analyse de la capacité germinative de quelques espèces d'acacia exotique. Tunis, Tunisie, Institut National de Recherches en Génie Rural Eaux et Forêts, 247p
- [61] Huang, J. and R. Redmann, 1995, Salt tolerance of Hordeum and Brassica species during germination and early seedling growth. Canadian Journal of Plant Science, 75(4), 815-819
- [62] Croser, C., S. Renault, J. Franklin and J. Zwiazek, 2001, The effect of salinity on the emergence and seedling growth of Picea mariana, Picea glauca, and Pinus banksiana. Environmental Pollution, 115(1), 9-16
- [63] Grieve, C. and L. Francois, 1992, The importance of initial seed size in wheat plant response to salinity. Plant and Soil, 147(2), 197-205
- [64] Soltani, A., M. Gholipoor and E. Zeinali, 2006, Seed reserve utilization and seedling growth of wheat as affected by drought and salinity. Environmental and Experimental Botany, 55(1–2), 195-200
- [65] Katembe, W.J., I.A. Ungar and J.P. Mitchell, 1998, Effect of salinity on germination and seedling growth of twoAtriplexspecies (Chenopodiaceae). Annals of Botany, 82(2), 167-175
- [66] Guan, B., D. Zhou, H. Zhang, Y. Tian, W. Japhet and P. Wang, 2009, Germination responses of Medicago ruthenica seeds to salinity, alkalinity, and temperature. Journal of Arid Environments, 73(1), 135-138
- [67] Chérifi, K., E. Boufous and A. El Mousadik, 2011, Diversity of Salt Tolerance During Germination in Medicago ciliaris (L.) and Medicago polymorpha (L.). Atlas Journal of Plant Biology, 1(1), 6-12
- [68] Khayatnezhad, M., R. Gholamin, S. Jamaati-e-Somarin and R. Zabihi-emahmoodabad, 2010, Study of NaCl salinity effect on wheat (Triticum aestivum L.) cultivars at germination stage. American-Eurasian J Agric Environ Sci, 9, 128-132
- [69] Tilaki, G.A.D., F. Gholami, K.G. Bezdi and B. Behtari, 2014, Germination percentage and recovery of Lolium perenne L. and Bromus tomentellus Boiss.(Poaceae, Liliopsida) seeds at several osmotic potential levels of iso-osmotic solutions. ПОВОЛЖСКИЙ ЭКОЛОГИЧЕСКИЙ ЖУРНАЛ, 2, 284-292
- [70] Pujol, J.A., J.F. Calvo and L. Ramirez-Diaz, 2000, Recovery of germination from different osmotic conditions by four halophytes from southeastern Spain. Annals of Botany, 85(2), 279-286
- [71] Ungar, I.A., 2001, Seed banks and seed population dynamics of halophytes. Wetlands Ecology and Management, 9(6), 499-510
- [72] Khan, M.A. and I.A. Ungar, 1984, Seed polymorphism and germination responses to salinity stress in Atriplex triangularis Willd. Botanical Gazette, 145(4), 487-494
- [73] Khayatnezhad, M. and R. Gholamin, 2011, Effects of water and salt stresses on germination and seedling growth in two durum wheat (Triticum durum Desf.) genotypes. Scientific Research and Essays, 6(21), 4597-4603
- [74] Prisco, J.T., J. Eneas Filho and E. Gomes Filho, 1981, Effect of NaCl salinity on cotyledon starch mobilization during germination of Vigna unguiculata (L.) Walp seeds. Revista brasileira de Botánica, 4, 63-71
- [75] El-Keblawy, A. and A. Al-Rawai, 2005, Effects of salinity, temperature and light on germination of invasive Prosopis juliflora (Sw.) DC. Journal of Arid Environments, 61(4), 555-565
- [76] Grabe, D.F., 1976, Measurement of seed vigor. Journal of Seed Technology, 1(2), 18-32
- [77] Khan, M.A. and M. Qaiser, 2006. Halophytes of Pakistan: characteristics, distribution and potential economic usages. Sabkha ecosystems, Springer: 129-153