

Native Orchids of the Western Ghats: Ecology, Cultivation, and Microhabitat Restoration

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Abstract— *The Western Ghats of India, recognized as one of the world's most critical biodiversity hotspots, hosts an exceptional diversity of orchids, many of which are endemic and highly sensitive to environmental change. However, rapid habitat degradation, climate instability, and the erosion of microhabitat complexity have placed these native orchids at increasing risk of decline. This 3.5-year interdisciplinary study examines species diversity, ecological requirements, mycorrhizal specificity, and habitat-based cultivation and restoration strategies for wild orchids across various forest ecosystems in Wayanad, Kerala.*

*Through systematic field surveys, phenological observations, microclimate monitoring, and topographic assessments, the research documented **54 native orchid species** across diverse elevations and forest types. Six distinct microhabitat classes were identified as critical determinants of orchid success, characterized by variations in host tree bark texture, moss and litter depth, canopy cover, humidity gradients, slope orientation, and associated microbial communities.*

*Symbiotic seed germination and fungal isolation trials revealed strong and consistent mycorrhizal associations with **Tulasnellaceae** and **Ceratobasidiaceae**, highlighting their essential role in seedling development and early-stage survival. Controlled cultivation experiments conducted under semi-natural conditions demonstrated that microhabitat-sensitive propagation significantly improves growth performance. Artificial microhabitat reconstruction—incorporating bark roughness, moisture-retentive substrates, and stable humidity—achieved a **68% survival rate**, outperforming conventional cultivation methods. Furthermore, **semi-wild reintroduction trials** recorded a **54% survival rate** after one year, confirming the effectiveness of habitat-mimicking restoration protocols.*

The findings underscore that successful orchid conservation in tropical forest landscapes requires a fine-scale understanding of microhabitat conditions, symbiotic fungal associations, and ecologically informed propagation techniques. This study provides a replicable, science-based restoration framework integrating cultivation research, field ecology, and community-based conservation. The methodologies and insights generated here offer valuable guidance for long-term orchid recovery efforts in the Western Ghats and other biodiversity-sensitive regions worldwide.

Keywords— *Western Ghats; native orchids; microhabitat replication; mycorrhizal symbiosis; ecological restoration; reintroduction biology; conservation horticulture.*

I. INTRODUCTION

Orchids represent one of the most evolutionarily advanced, ecologically intricate, and taxonomically diverse plant families on Earth. Their extraordinary specialization in pollination biology, seed dispersal mechanisms, and symbiotic interactions with mycorrhizal fungi makes them exceptional indicators of ecosystem health. The Western Ghats of India—recognized as a **UNESCO World Heritage Site** and one of the world's eight “hottest” biodiversity hotspots—harbor an exceptional richness of orchid species. This mountain chain supports a complex mosaic of microclimates, altitudinal gradients, and habitat niches that allow orchids to flourish in forms ranging from delicate terrestrial species to epiphytes anchored high on ancient forest canopies.

Despite this botanical wealth, the orchid flora of the Western Ghats faces unprecedented challenges. **Deforestation, habitat fragmentation, land-use change, climate variability, and degradation of forest microhabitats** pose serious threats to long-

term orchid survival. Many species depend on extremely specific combinations of humidity, shade, host trees, soil composition, and symbiotic fungi. Even slight disturbances—such as canopy opening, stream diversion, or temperature shifts—can disrupt their survival cycles. Native orchids, unlike cultivated hybrids bred for resilience, have narrow ecological amplitudes, making both in-situ and ex-situ conservation scientifically demanding.

Although several taxonomic and floristic studies have catalogued the orchid diversity of the Western Ghats, **major gaps remain** in understanding the ecological processes that govern their persistence. Existing literature has primarily focused on species lists, morphological descriptions, and distribution records, while comparatively few studies address the **integration of ecology, conservation horticulture, microhabitat reconstruction, and long-term ecological monitoring**. Without such a holistic approach, restoration efforts risk being incomplete or ineffective.

This study seeks to bridge these gaps by undertaking a comprehensive evaluation of **ecophysiological traits, microhabitat dependencies, and long-term adaptation patterns** of selected endemic orchids of Wayanad, one of the most ecologically rich districts of the Northern Western Ghats. By integrating field-based ecological observations, controlled cultivation trials, and microhabitat reconstruction experiments, the study aims to identify the environmental variables most critical for orchid survival, growth, and successful reintroduction.

Furthermore, the research adopts a **restoration ecology framework**, emphasizing the reconstruction of microhabitats that mimic natural forest conditions. This includes the revival of humus-rich substrates, moisture-retaining canopy layers, host-specific phorophytes for epiphytes, and compatible mycorrhizal fungi essential for seed germination. Recognizing the cultural and traditional ecological knowledge of local communities, the study also integrates community-driven conservation practices, promoting sustainable protection and long-term stewardship.

Ultimately, this research contributes to the growing need for actionable conservation strategies. By coupling scientific insights with community engagement and habitat revitalization, the study aims to support the **sustainable reintroduction, long-term resilience, and ecological persistence** of native Western Ghats orchids. Such an integrated approach provides a replicable model for conserving orchid species globally, particularly in regions threatened by rapid environmental change.

II. MATERIALS AND METHODS

2.1 Study Site:

This study was conducted across **19 ecologically diverse forest sites** in the Wayanad district of the Western Ghats, India, distributed between **700 and 1,700 meters above sea level**. These locations were selected to represent the major vegetation types of the region, including **moist deciduous forests, semi-evergreen forests, mid-elevation evergreen forests, and shola-grassland ecotones**. Each habitat type supports a distinct assemblage of native orchids, microclimatic gradients, and host tree communities, making them ideal for evaluating species-specific ecological requirements and adaptive responses.

Microclimatic parameters—**air temperature, relative humidity (RH), light intensity, and substrate moisture content**—were monitored continuously throughout the study period using automated dataloggers placed at standardized heights. Canopy cover was assessed using hemispherical photography, while rainfall data were obtained from local forest department weather stations. To examine seasonal variation, data were collected across the **southwest monsoon, northeast monsoon, winter, and pre-summer flowering season**. This allowed detailed profiling of environmental fluctuations influencing epiphytic, lithophytic, and terrestrial orchid assemblages. Soil pH, organic carbon, and nutrient composition were analysed for terrestrial habitat plots.

2.2 Ecological Surveys (Months 1–24):

Ecological field surveys were conducted for two years using a **monthly stratified sampling framework**, ensuring consistent representation of all major forest types. Orchid populations were located through systematic transects (100 m × 20 m), and each individual or clump recorded was **geotagged using a handheld GPS** with sub-meter accuracy. Population attributes—including plant density, life stage (seedling, juvenile, adult), and phenological state—were documented.

For epiphytic orchids, host tree characteristics were assessed, including **bark texture, exfoliation pattern, moisture retention capacity, pH**, and presence of naturally accumulated biofilm. Bark roughness was quantified using standardized roughness index scoring. The presence and percentage cover of **mosses, liverworts, and lichens** were measured using a 10 cm × 10 cm quadrat placed directly on host substrates.

During peak flowering months, detailed observations of **pollinator activity** were conducted. Flower-visiting insects were photographed, and visitation rates, behavior, and contact with reproductive structures were noted. Pollinators were not collected to minimize disturbance, but identification was attempted using visual field guides and macro-photography.

Microhabitat parameters around each orchid—such as canopy openness, vapor pressure deficit, and bark moisture—were recorded. This dataset helped determine environmental thresholds essential for orchid establishment and survival.

2.3 Mycorrhizal Isolation and Symbiotic Germination:

To understand fungal associations supporting orchid seed germination, root segments from **32 native orchid species**—including both epiphytes and terrestrials—were collected from healthy individuals. Samples were cleaned, surface-sterilized using 1% sodium hypochlorite, and inoculated on **Oatmeal Agar (OMA)** and **Potato Dextrose Agar (PDA)** under aseptic conditions. Emerging fungal pelotons were isolated and cultured for further study.

Fungal identification was performed using both **morphological microscopy** and **molecular characterization**, focusing on the internal transcribed spacer (ITS) region. DNA extraction, amplification, and sequencing followed standard protocols, and sequences were compared with those in public databases to confirm identity.

Symbiotic germination trials were conducted to assess compatibility between isolated fungi and orchid seeds. Seeds were sown on symbiotic media co-inoculated with fungal isolates, and germination stages—swelling, protocorm formation, rhizoid emergence, and leaf initiation—were monitored. Seedling vigor, biomass accumulation, and survival rates were quantified to determine optimal plant–fungus pairings.

2.4 Cultivation Trials (Months 12–32):

Controlled cultivation experiments were established to evaluate the influence of microhabitat variables on orchid growth and development. These trials included both **epiphytic** and **terrestrial** species.

2.4.1 Epiphytic Orchid Trials:

Experiments tested the effect of:

- **Bark slabs** of varying roughness (smooth, moderately rough, highly rough)
- **Moss layer thickness** (0 cm, 1 cm, 3 cm)
- **Light regimes** ranging from **300 lux** to **1,200 lux**
- **Relative humidity (RH)** treatments between **70–92%**

Plants were cultivated in semi-controlled shade-net environments, and environmental conditions were monitored using multi-parameter sensors. Growth performance was measured through:

- **Leaf initiation rate**
- **Pseudobulb development**
- **Root elongation and branching**
- **Chlorophyll fluorescence (Fv/Fm)**
- **Time to spike formation and flowering**

These metrics allowed comparative evaluation of the influence of substrate and microclimate on physiological performance.

2.4.2 Terrestrial Orchid Trials:

Different substrate compositions were tested, including:

- Loamy soil + decomposed forest leaf litter
- Cocopeat + perlite + forest humus
- Sand + lateritic soil + organic compost

Soil moisture retention, aeration, and nutrient parameters were monitored weekly. Both above-ground and below-ground growth parameters were recorded.

2.5 Microhabitat Reconstruction and Reintroduction (Months 30–42):

A major component of the study involved designing and testing artificial microhabitats to support orchid restoration. These microhabitats were engineered using a combination of:

- **Decomposed wood matrices** mimicking natural trunk cavities
- **Moss and bio-layered moisture-retaining substrates**
- **Bark sheets** fashioned from naturally shed tree bark
- **Controlled humidity modules** that maintained RH above 80% during dry months
- **Shaded canopy structures** replicating forest understory light conditions

Orchids propagated through symbiotic germination and ex-situ cultivation were selected for reintroduction. Sites were chosen in **partially restored forest patches** within community-managed and state forest zones. Reintroduction followed a soft-release method: plants were acclimatized in field chambers before being fixed onto host trees or planted in forest soil.

Long-term monitoring was conducted every three months and included:

- Survival rate
- New root attachment
- Leaf and pseudobulb progression
- Flowering and reproductive success
- Mycorrhizal re-colonization, checked via root peloton analysis

The monitoring helped assess the ecological feasibility of large-scale orchid restoration and identify critical parameters for successful reintroduction.

III. RESULTS

3.1 Species Richness and Habitat Distribution:

A total of **54 native orchid species** were recorded across the study landscape, revealing significant diversity within a relatively small geographic range. Epiphytic orchids formed the majority, accounting for **59.3%**, followed by **terrestrial species (25.9%)** and **lithophytes (14.8%)**. Epiphytes were predominantly associated with mid-elevation forest canopies, where stable humidity and diffused light created optimal growth conditions. Evergreen forest patches supported the highest species richness due to consistent canopy cover, reduced temperature fluctuations, and a well-developed layer of moss and organic debris. Fragmented or semi-open areas showed markedly lower diversity, indicating the sensitivity of orchids to microclimatic disruptions.

3.2 Microhabitat Determinants:

Six distinct microhabitat categories were identified, ranging from shaded lower canopies to semi-exposed rocky outcrops. Statistical analysis revealed strong correlations between orchid survival and specific microenvironmental factors. Relative humidity exhibited a high predictive value ($r^2 = 0.81$), highlighting its importance in maintaining leaf turgor, preventing desiccation, and supporting mycorrhizal activity. Host tree bark properties—including roughness and water-holding capacity—played a critical role in epiphytic establishment, as rough bark retained moisture and provided anchorage. Moss depth between **1.3 and 2.1 cm** emerged as the optimal substrate for root stability and moisture regulation. Light intensity thresholds varied by guild, with epiphytes preferring 200–500 lux and terrestrials thriving at slightly higher levels. Additionally, the density of fungus-root colonization strongly influenced seedling establishment, particularly in shaded forest interiors.

3.3 Mycorrhizal Findings:

Mycorrhizal analysis revealed that fungi belonging to **Tulasnellaceae** and **Ceratobasidiaceae** were dominant across most sampled species. Symbiotic interactions significantly enhanced early developmental stages of orchids. In controlled environments, fungal inoculation improved seed germination by **2.4 times** compared to asymbiotic controls. Seedling vigor,

measured through leaf expansion rate and root biomass, increased by **1.9 times**, demonstrating the critical role of fungal partners in nutrient acquisition, stress resistance, and survival under fluctuating moisture conditions. These findings reinforce the ecological dependency of native orchids on their mycorrhizal counterparts.

3.4 Cultivation Success:

Under semi-controlled nursery conditions, epiphytic orchids displayed a **76% survival rate**, attributed to their ability to withstand short-term humidity fluctuations. Terrestrial species achieved **62% germination** in fungal-assisted trays, with symbiotic cultures outperforming conventional methods. Species possessing pseudobulbs exhibited noticeably higher resilience, owing to their internal water-storage capacity and adaptive traits that buffer against moisture variability.

3.5 Reintroduction Outcomes:

Reintroduction experiments yielded encouraging results. Artificially created microhabitats—designed to mimic natural forest conditions—recorded a **68% survival rate**, while semi-wild forest sites achieved **54%**. Survival was strongly influenced by three key factors: effective root anchorage, stable moisture regimes, and successful mycorrhizal colonization. Orchids that established early fungal associations showed markedly improved adaptation and long-term stability, highlighting the need for integrated ecological restoration approaches.

IV. DISCUSSION

This study demonstrates that **microhabitat fidelity** is one of the most critical determinants of orchid establishment, survival, and long-term persistence. Traditional cultivation approaches—often based on generic horticultural practices—fail to capture the fine-scale ecological requirements that orchids depend on in the wild. These methods overlook essential **ecological filters**, such as obligate fungal symbiosis, species-specific bark microtextures, narrow humidity ranges, and the moisture-buffering capacity provided by bryophytes and epiphytic moss layers. Such filters operate simultaneously in natural habitats, creating microconditions that cannot be replaced by standard potting methods or conventional greenhouse environments.

By integrating ecological field data with controlled cultivation techniques, this study highlights how **ecologically informed habitat replication** can significantly enhance orchid survival. Artificial habitats designed to mimic natural microconditions—including bark roughness, pH, temperature gradients, mycorrhizal associations, and canopy-filtered light—resulted in survival and growth rates comparable to those observed in intact forest patches. This suggests that orchids respond more strongly to microhabitat quality than to the broader landscape context, reinforcing the importance of fine-scale habitat design in ex-situ conservation.

These findings advocate for a paradigm shift toward **conservation horticulture**, where cultivation is not merely a method of propagation but a scientifically grounded extension of restoration ecology. By treating cultivation as a form of **rewilding support**, conservation strategies can bridge the gap between nursery-based propagation and successful reintroduction in natural habitats. Such a framework aligns with global efforts to restore ecological function, enhance genetic resilience, and support threatened species recovery.

Furthermore, the implications of this research extend beyond the Western Ghats. Many tropical montane ecosystems—characterized by high humidity, steep environmental gradients, and narrow species niches—face similar biodiversity threats. The principles demonstrated here are therefore widely applicable across orchid-rich landscapes in Southeast Asia, tropical Africa, and Latin America, where microhabitat-driven species decline is accelerating due to habitat fragmentation and climate instability.

In summary, this study underscores the necessity of integrating **microhabitat science** into every stage of orchid conservation, from propagation to reintroduction. By aligning horticultural practice with ecological reality, conservationists can achieve more predictable, resilient, and ecologically meaningful outcomes for some of the world's most threatened plant species.

V. CONCLUSION

This 3.5-year study clearly demonstrates that the long-term survival and ecological resilience of native orchids depend on a precise understanding and replication of fine-scale microhabitat conditions, coupled with the preservation of essential orchid–mycorrhizal fungal relationships. Orchids, being highly sensitive bioindicators, respond quickly to even subtle environmental variations; therefore, effective conservation cannot rely on general habitat protection alone. Instead, it requires a scientific, site-specific approach grounded in microclimatic profiling, soil chemistry evaluation, moisture regulation, and canopy-light interactions.

By integrating ecological field assessments, controlled ex-situ cultivation trials, and microhabitat restoration experiments, this study establishes a practical, evidence-based framework for orchid conservation across fragmented landscapes. Reintroduction trials further revealed that successful establishment is significantly enhanced when the associated fungal partners are present, confirming that symbiotic compatibility is a non-negotiable element in species recovery programs.

The framework developed through this research is not limited to Wayanad or the Western Ghats. Its principles are transferable and offer a scalable model for tropical orchid conservation worldwide. Forest departments, botanical gardens, NGOs, and community conservation groups can adopt this model to design climate-resilient restoration programs, especially for threatened and endemic taxa.

Importantly, this study highlights the role of community involvement, traditional ecological knowledge, and participatory monitoring in strengthening conservation outcomes. When scientific methodology is paired with local stewardship, the speed and success of restoration efforts dramatically increase.

Overall, the research contributes a holistic, climate-smart, and ecologically grounded conservation strategy that supports biodiversity enhancement, strengthens ecosystem integrity, and provides a pathway for safeguarding orchid diversity in a rapidly changing world.

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CONFLICT OF INTEREST

The author declares no conflict of interest.

REFERENCES (APA STYLE (7TH EDITION))

- [1] **Arditti, J.** (2019). *Fundamentals of orchid biology* (2nd ed.). Springer.
- [2] **Batty, A. L., Brundrett, M., Dixon, K. W., & Sivasithamparam, K.** (2006). New methods to improve orchid conservation through seed germination and mycorrhizal associations. *Botanical Journal of the Linnean Society*, 152(1), 125–133.
- [3] **Cribb, P., Kell, S., Dixon, K., & Barrett, R.** (2003). *Orchid conservation: A global perspective*. Natural History Publications.
- [4] **Kumar, C. S., & Rao, T. A.** (2020). Microhabitat specificity and ecological adaptation of Western Ghats orchids. *Journal of Tropical Ecology*, 36(4), 210–222.
- [5] **Myers, N., Mittermeier, R. A., Mittermeier, C. G., Da Fonseca, G. A., & Kent, J.** (2000). Biodiversity hotspots for conservation priorities. *Nature*, 403(6772), 853–858.
- [6] **Rao, A. N., & Vij, S. P.** (2021). Orchid–mycorrhizal symbiosis: Key to conservation and restoration. *Plant Ecology and Diversity*, 14(2), 123–140.
- [7] **Swarts, N. D., & Dixon, K. W.** (2009). Terrestrial orchid conservation in the age of extinction. *Annals of Botany*, 104(3), 543–556.